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(54) Title: MRI IMAGE ENHANCEMENT OF BONE AND RELATED TISSUE USING COMPLEXES OF PARAMAGNETIC CATIONS AND POLYPHOSPHONATE LIGANDS

(57) Abstract

Polyphosphonate ligands containing three or more phosphonate groups, combined with paramagnetic metal cations and administered in the form of pharmacologically acceptable salts, are useful as MRI contrast enhancement agents, which tend to localize in bone tissue without being conjugated to bone-specific biomolecules. Triazacyclononanes and tetraazacyclododecanes, with dihydroxyphosphorylmethyl or dihydroxyphosphorylethyl groups, optionally substituted at the methyl or ethyl bridges with alkyl, aryl, hydroxyl or amino groups, are particularly preferred.

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MRI IMAGE ENHANCEMENT OF BONE AND RELATED TISSUE
USING COMPLEXES OF
PARAMAGNETIC CATIONS AND POLYPHOSPHONATE LIGANDS

5 This invention lies in the field of magnetic resonance imaging, and is relevant to the art of contrast enhancement agents used in connection with magnetic resonance imaging in medical diagnostics.

BACKGROUND OF THE INVENTION

10 The availability of magnetic resonance imaging (MRI) devices has led to the use of MRI in medical examinations for the detection and diagnosis of disease states and other internal abnormalities. The continued use and development of MRI has stimulated interest in the
15 development of pharmaceutical agents capable of altering MRI images in diagnostically useful ways. Pharmaceutical agents (MRI pharmaceuticals) which are currently favored by
20 researchers in the field are suitably complexed paramagnetic metal cations. The use of pharmaceuticals in MRI imaging offers major opportunities for improving the value of the diagnostic information which can be obtained

Radiopharmaceuticals, which are used in radioisotopic imaging in a manner analogous to MRI pharmaceuticals, are a well developed field. The knowledge
25 existing in this field thus provides a starting point for the development of MRI pharmaceuticals. MRI pharmaceuticals must meet certain characteristics, however, which are either not required or are considerably less critical in the case
30 of radiopharmaceuticals. MRI pharmaceuticals must be used in greater quantities than radiopharmaceuticals. As a result, they must not only produce detectable changes in proton relaxation rates, usually expressed as proton longitudinal relaxivity or $1/T_1$, but they must also be (a) substantially less toxic, thereby permitting the use of

greater amounts, (b) more water soluble to permit the administration of a higher dosage in physiologically acceptable volumes of solution, and (c) more stable in vivo than their radiopharmaceutical counterparts. In vivo stability is important in preventing the release of free paramagnetic metals and free ligand in the body of the patient, and is likewise more critical due to the higher quantities used. For the same reasons, MRI pharmaceuticals which exhibit whole body clearance within relatively short time periods are particularly desirable.

Since radiopharmaceuticals are administered in very small dosages, there has been little need to minimize the toxicity of these agents while maximizing water solubility, in vivo stability and whole body clearance. It is not surprising therefore that few of the ligands developed for use as components in radiopharmaceutical preparations are suitable for use in preparation of MRI pharmaceuticals. A notable exception is the well known ligand diethylene triamine pentaacetic acid (DTPA), which has proved useful in forming complexes with both radiocations, pharmacologically suitable salts of which provided useful radiopharmaceuticals, and paramagnetic cations such as gadolinium, whose pharmacologically suitable salts have proved useful as MRI pharmaceuticals.

Certain groups of radiopharmaceuticals tend to localize in bone tissue, and are thus of use in providing diagnostic information concerning bone disorders. The properties of these agents which lead to their localization in bone also allow for them to be used to gain useful information concerning the function of the kidneys, the size and location of myocardial infarcts, regional breakdowns in the blood brain barrier, and other organic disorders similarly related. It is clear that employment of bone localizing paramagnetic agents in MRI imaging would provide similarly specific diagnostic information. Unfortunately, no MRI agents which have this characteristic have been described.

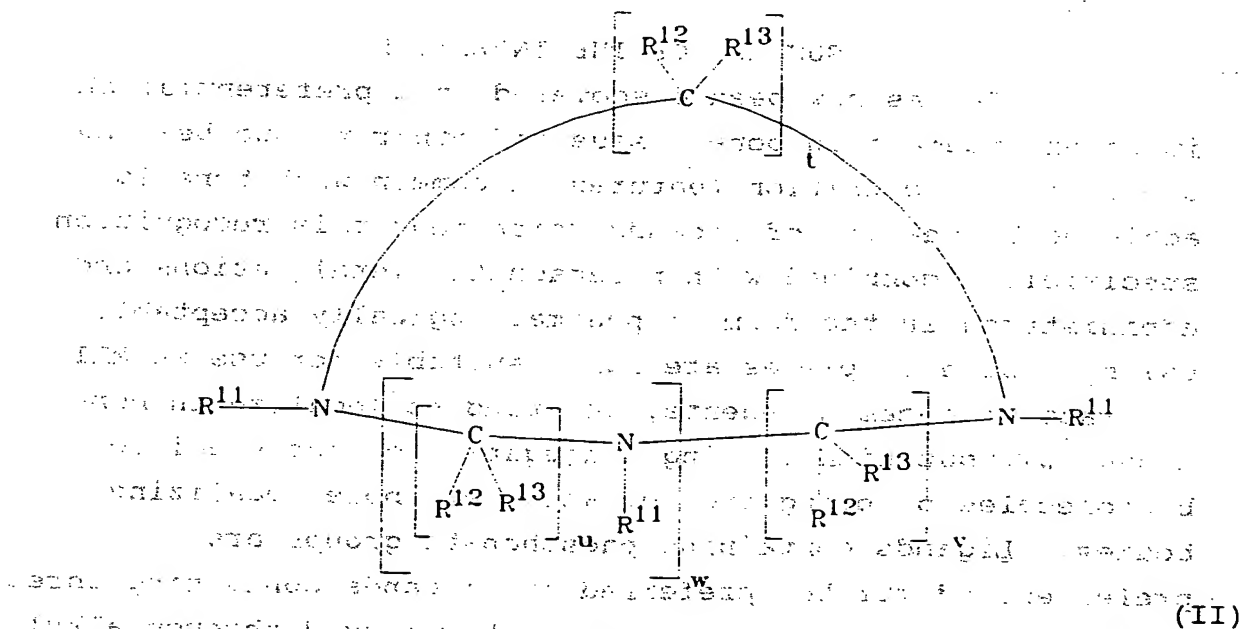
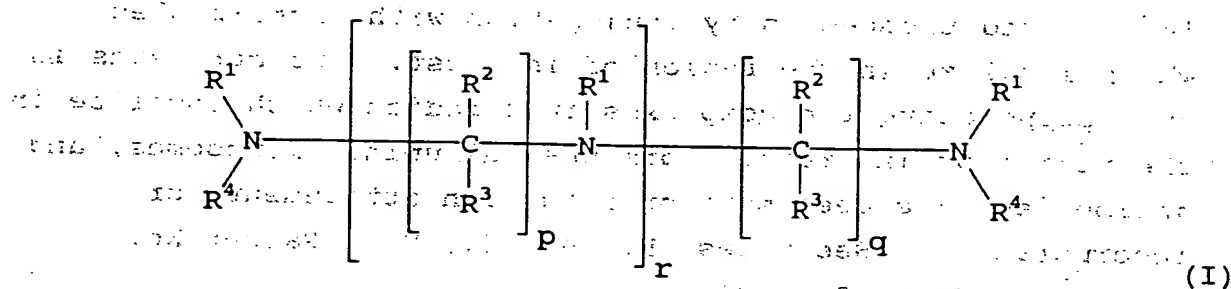
Most known MRI pharmaceuticals when administered in vivo do not by themselves localize in specific tissues, but instead generally distribute in extracellular fluid space in a nonspecific manner. One means of achieving localization of these inherently nonspecific pharmaceuticals in selected tissues is by conjugation with biomolecules which localize in the region of interest. Another means is by incorporating the complexes into bodies which localize in the region of interest. Hormones, albumins, liposomes, and antibodies have been mentioned in such attachments or incorporation. [See Gries, H., et al., U.S. Patent No. 4,647,447, March 3, 1987.

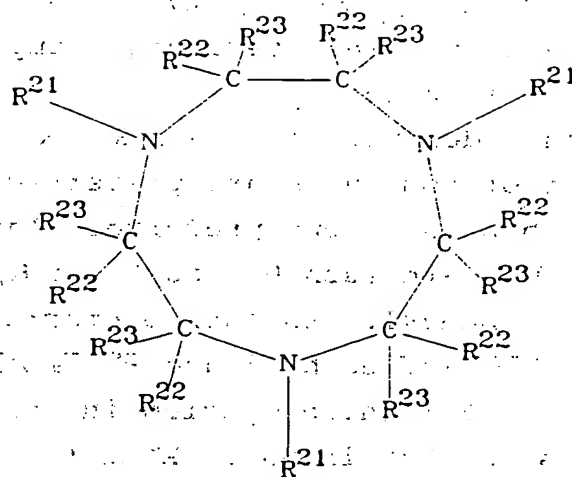
SUMMARY OF THE INVENTION

It has now been discovered that preferential MRI image enhancement in bone tissue and other tissue bearing biospecific recognition features in common with bone is achieved by the use of ligands which bear this recognition specificity, combined with paramagnetic metal cations and administered in the form of pharmacologically acceptable salts. These complexes are fully suitable for use as MRI contrast enhancement agents, and tend to localize in bone tissue without either being conjugated to bone-specific biomolecules or being incorporated into bone localizing bodies. Ligands containing phosphonate groups are preferred, and further preferred are ligands containing three or more phosphonate groups, preferably bonded through alkyl bridges to nitrogen atoms. Cyclic groups are still further preferred, notably polyazacycloalkanes. Particularly preferred ligands are triazacyclononanes and tetraazacyclododecanes, with dihydroxyphosphorylmethyl or dihydroxyphosphorylethyl groups attached to the nitrogen atoms, these groups optionally substituted at the methyl or ethyl bridges with alkyl, aryl, hydroxyl or amino groups.

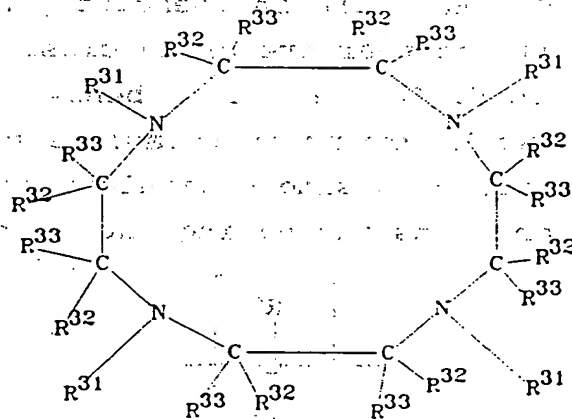
DETAILED DESCRIPTION OF THE INVENTION AND PREFERRED EMBODIMENTS

Among the ligands used in the practice of the present invention are the embodiments represented by the following formulas:



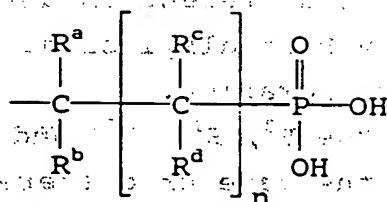


(III)



(IV)

The R¹, R⁴, R¹¹, R²¹ and R³¹ groups in these formulas are phosphonate groups which may be the same or different on any particular species, and are generally represented by



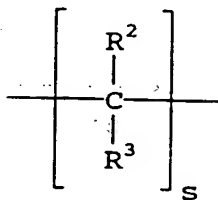
in which:

R^a, R^b and R^c are independently H, or alkyl or aryl groups which do not interfere with complexation;

R^d is H, OH, NH_2 , or alkyl or aryl groups which do not interfere with complexation; and n is zero or 1.

In this definition of R^1 , R^4 , R^{11} , R^{21} , and R^{31} , certain classes of compounds are preferred. For those species in which n is 1, one preferred class is that in which R^a , R^b and R^c are each H; and R^d is H, OH, NH_2 , C_1-C_8 alkyl, phenyl or benzyl. Another preferred class is that in which R^a , R^b and R^c are each H; and R^d is H, OH, NH_2 , C_1-C_4 alkyl or benzyl. For those species in which n is zero, a preferred class is that in which R^a and R^b are independently H, C_1-C_4 alkyl or benzyl, while another preferred class is that in which R^a and R^b are independently H, C_1-C_4 alkyl or benzyl, and still another preferred class is that in which R^a is H and R^b is H, C_1-C_4 alkyl or benzyl.

The two R^4 groups in Formula I may alternatively be joined together as a single divalent group bridging the two end nitrogen atoms and having the formula



in which R^2 and R^3 are as defined below, and s is at least 2, preferably 2 or 3.

The R^2 , R^{12} , R^{22} and R^{32} groups in these formulas may also be the same or different on any single species, and are each independently H or alkyl or aryl groups which do not interfere with complexation.

Similarly, the R^3 , R^{13} , R^{23} and R^{33} groups in these formulas may also be the same or different on any single species, and are each independently H or alkyl or aryl groups which do not interfere with complexation.

In Formula I, the subscripts p and q may be the same or different, and are each either 2 or 3. The subscript r is 0 to 3 inclusive, preferably 0 to 2 inclusive, and most preferably 0 or 1.

In Formula II, t, u and v may be the same or different, and are each either 2 or 3. The value of w is at least 1, more preferably 1 to 4 inclusive, still more preferably 1 to 3 inclusive, and most preferably either 1 or 2.

In preferred embodiments, all R^1 , R^{11} , R^{21} or R^{31} groups on any single species are the same. In further preferred embodiments, all R^2 , R^{12} , R^{22} or R^{32} groups on any single species are the same, and all R^3 , R^{13} , R^{23} or R^{33} groups on any single species are the same. In still further preferred embodiments, all R^2 , R^{12} , R^{22} or R^{32} groups on any single species are H, and all R^3 , R^{13} , R^{23} or R^{33} groups on any single species are the same and are H or alkyl or aryl groups which do not interfere with complexation. In still further preferred embodiments, all R^2 , R^{12} , R^{22} or R^{32} groups as well as all R^3 , R^{13} , R^{23} or R^{33} groups on any single species are H.

The complexation referred to in the descriptions of the alkyl and aryl groups is the complexation of the ligand with a paramagnetic metal cation to form a chelate. Alkyl and aryl groups which do not interfere with such complexation extend to a wide range in terms of size and configuration. Preferred alkyl groups are those having 1 to 8 carbon atoms, with 1 to 4 carbon atom alkyls more preferred, and methyl and ethyl the most preferred. Both straight-chain and branched-chain alkyls are included. Preferred aryl groups are benzyl and phenyl, particularly benzyl.

Paramagnetic metals of a wide range are suitable for complexation with these ligands in the formation of the contrast enhancement agents of the present invention. These metals tend to focus in elements having atomic numbers of 22-29 (inclusive), 42, 44 and 58-70 (inclusive), and have oxidation states of 2 or 3. Of these, the ones having atomic number of 22-29 (inclusive) and 58-70 (inclusive) are preferred, and those having atomic numbers of 24-29 (inclusive) and 64-68 (inclusive) are more preferred.

Examples of such metals are chromium (III), manganese (II), iron (II), iron (III), cobalt (II), nickel (II), copper (II), praseodymium (III), neodymium (III), samarium (III), gadolinium (III), terbium (III), dysprosium (III), holmium (III), erbium (III) and ytterbium (III). Chromium (III), manganese (II), iron (III) and gadolinium (III) are particularly preferred, with iron (III) the most preferred.

Physiologically or pharmacologically compatible salts of the chelates are formed by neutralizing acidic

moieties on the chelate with physiologically or pharmacologically compatible cations from corresponding inorganic and organic bases and amino acids. Examples include alkali and alkaline earth metal cations, notably sodium. Further examples are primary, secondary and tertiary amines, notably, ethanolamine, diethanolamine, morpholine, glucamine, N,N-dimethylglucamine, and N-methylglucamine (commonly referred to as "meglumine").

Examples of amino acid cations are lysines, arginines and ornithines. As bases, these cations may be used in the form of hydroxides, carbonates, bicarbonates or any other base forms which will release the cations. Of the many embodiments of the present invention, one preferred class consists of the physiologically compatible salts which contain three equivalents of a physiologically compatible cation combined with the trianionic complex of Fe(III) and N,N',N''-tris(dihydroxyphosphorylmethyl)-1,4,7-triazacyclononane at a pH of 6.8 to 7.4. (The term "trianionic" in this context denotes an anion having a charge of -3.)

The compounds of the present invention are capable of preparation by known procedures, some of which are described herein. The phosphonic acid, referred to herein as the "ligand," is first formed, followed by the formation of the chelate complex and then the physiologically compatible salt.

According to a typical procedure, compounds with a methylene bridge between the N and P atoms (i.e., those in which n in the above formulae is zero) are prepared by first

treating the hydrobromide salt of the unsubstituted starting material (for example, 1,4,7-triazacyclononane or 1,4,7,10-tetraazacyclododecane) with formaldehyde and diethyl phosphite in aqueous solution to form the perethyl phosphonate ester (i.e., all acid groups esterified with an ethyl group). The ester subsequently can be hydrolyzed to the phosphonic acid ligand. Alkyl or aryl substitutions are introduced on the methylene carbon by treatment of the perethyl ester with a strong base such as butyllithium at -78°C and an alkyl or aryl halide.

Likewise, the preparation of compounds with an ethylene bridge between the N and P atoms (n equaling 1) from the same unsubstituted starting materials is begun by treating the starting materials with diethyl 2-bromoethylphosphonate in the presence of excess K_2CO_3 . This will form the phosphonic acid perethyl esters, which are then hydrolyzed in the same manner as the methylene bridge compounds.

Ethylene bridge compounds with a hydroxy substitution at the carbon adjacent to the P atom (i.e., as R^d) are prepared by using diethyl epoxyethylphosphonate in place of the diethyl 2-bromoethylphosphonate, and base is not used in the reaction. Those skilled in the art will recognize that similar compounds containing an amino substitution in the position of the hydroxy substitution can be prepared similarly using diethyl ethyleniminophosphonate.

It was discovered that the procedure for combining the ligand with a paramagnetic metal cation to form the chelate complex is critical when seeking to obtain a stable, chromatographically distinct species. In particular, for most of the complexes studied it was discovered that a stable distinct species was obtained by heating a solution of the ligand and a water soluble compound of the metal cation to a temperature of at least about 50°C, preferably at least about 80°C, and more preferably to reflux (100°C in an aqueous system), at a pH in excess of 7.0. In preferred embodiments, separation and purification are incorporated

into the process of elevation of the pH and heating. Thus, after initially adding the acid form of the ligand and the halide form of the paramagnetic cation and heating, the pH is slowly elevated by slow addition of base in an amount of 5 equivalents equal to the charge of the metal cation. Thus, when the metal cation is Mn(II), two equivalents of base are added, and when the cation is Fe(III), three equivalents are added. The neutral form of the complex can then usually be crystallized as a solid from the solvent. While heating, 10 the crystallized solid can be added to water and sufficient base to neutralize all remaining labile protonated sites of the complex. Following formation of the chromatographically distinct complex, the neutral form of the complex can then typically be recrystallized following reacidification. The 15 optimum temperature and base addition rate will vary from one metal ion to the next, and is readily determined by routine experimentation. In certain cases, (e.g., the Fe(III) complex of N,N',N''-tris(dihydroxyphosphorylethyl)-1,4,7-triazacyclononane where the complex forms multiple 20 species on acidification), crystallization of the neutral complex from acid medium was not performed, and the desired salt was obtained directly from solution.

Use of the procedure described typically results in species which are stable against degradation into 25 multiple, chromatographically distinct species over time, and upon exposure to elevated temperature. The term "chromatographically distinct" is used herein to denote species which do not indicate separation into components 30 when subjected to suitable chromatography.

Any water soluble form of the metal may be used. Notable examples are halide salts and carbonate salts. Chlorides are particularly preferred. Sparingly water soluble oxides may also be used. When oxides are used, addition of base is not needed to form the neutral form of 35 the complex.

Physiological salts are prepared from the neutral forms of the complexes by conventional procedures. In a

typical procedure, the desired salt of the complex is formed from the neutral form of the complex by addition of the required equivalent of the desired base. Heating until the pH stabilizes may be required. A solid form of the salt of the complex can be obtained by conventional procedures, such as, for example, lyophilization, and the solid can be reconstituted with pharmacologically suitable aqueous solutions prior to administration to patients. The number of physiological cations present in the final product is equal to the equivalents added during the step of base addition and is readily confirmed by independent means such as elemental analysis or potentiometric titrations.

Administration of the MRI contrast agents of the present invention to a patient or subject on whom magnetic resonance imaging is to be performed is achieved by conventional procedures known in this art and disclosed in the literature. Aqueous solutions of the agents are most conveniently used. The concentrations of the agents in these solutions and the amounts administered may vary widely, the optimum in each case varying with the strength of the magnetic moment of the paramagnetic metal in the agent, the contrast enhancement strength of the chelate as a whole, the method of administration, the degree of contrast enhancement desired or needed, and the age, weight and condition of the patient or subject to whom administration is made. In most cases, best results are obtained with solutions at concentrations of about 0.05 to about 2.0 moles of the paramagnetic complex per liter, preferably about 0.1 to about 1.0 mole per liter. Likewise, best results in most cases are usually obtained with dosages ranging from about 0.01 to about 1.0 millimole of agent per kilogram of whole body weight (mM/kg), preferably from about 0.05 to about 0.5 mM/kg. Administration may be achieved by any parenteral route and method, most notably by intravenous administration. The rate of administration may likewise vary, best results generally being obtained at rates ranging from about 0.1 mM/min/kg to about 1.0 mM/sec/kg.

The following examples are offered for purposes of illustration, and are intended neither to define nor limit the invention in any manner.

EXAMPLE 1

SYNTHESES OF DIHYDROXYPHOSPHORYLMETHYL SPECIES

This example illustrates the preparation of various dihydroxyphosphorylmethyl compounds and complexes within the scope of the invention. Species based on both 1,4,7-triazacyclononane and 1,4,7,10-tetraazacyclododecane are illustrated in parallel fashion starting from the hydrobromide salts of 1,4,7-triazacyclononane and 1,4,7,10-tetraazacyclododecane, respectively.

A. Synthesis of Perethyl Esters of Nitrogen-

Substituted Methylene Phosphonates 1,4,7-Triaza-

cyclononane and 1,4,7,10-Tetraazacyclododecane

The trihydrobromide salt of 1,4,7-triazacyclononane and the hydrobromide salt of 1,4,7,10-tetraazacyclododecane were combined with 3.5 equivalents, and 28 equivalents, respectively, of aqueous 37% formaldehyde solution. The mixtures were stirred for 15-30 minutes at room temperature, after which time 3.5 equivalents and 14 equivalents, respectively, of diethyl phosphite were added to each solution, and the reaction mixtures were stirred at room temperature for an additional 2-5 hours. Water was

then added and the aqueous layers extracted five times with ethyl acetate. To the remaining water fractions, NaHCO_3 was added until the solutions were of pH approximately 7.5. The solutions were then continuously extracted with ether for 2-2.5 days. The products were obtained as oils upon evaporation of the ether and as needed were purified by chromatography, and were identified as the perethyl esters

of 1,4,7-triazacyclononane and 1,4,7,10-tetraazacyclododecane, respectively, by NMR.

Those skilled in the art will recognize that this procedure can also be employed to synthesize compounds

5 derived from substituted forms of 1,4,7-triazacyclononane and 1,4,7,10-tetraazacyclododecane where such substitutions are on the ring carbons and consist of alkyl or aryl groups as listed above, retaining the substitutions in the

corresponding positions on the product compounds. Those

10 skilled in the art will further recognize that this

procedure can be employed in an analogous manner to

synthesize other substituted and unsubstituted cyclical and linear polyamines.

15 B. Synthesis of Perethyl Esters of Nitrogen-

Substituted Methylene Phosphonates Substituted with Benzyl Groups at the Methylene Carbon

In this procedure, one of the perethyl esters

20 prepared in part A above is converted to an analog which contains a benzyl group attached to the methylene carbon.

The perethyl ester of 1,4,7-triazacyclononane prepared in part A above, dissolved in dry tetrahydrofuran, was combined with an excess of butyllithium at -78°C, and
25 the reaction mixture was stirred for 30 minutes at that temperature. An amount of benzyl bromide equal to the number of equivalents of butyllithium employed was then added with stirring. The mixture was then allowed to slowly warm to room temperature. After continued stirring at room
30 temperature for an additional 30 minutes, cold water was added and the aqueous layer was extracted with diethyl

ether. The ether was evaporated and the residue chromatographed on silica gel G60 70-230 mesh to obtain the perethyl ester of N,N',N''-tris(dihydroxyphosphorylbenzylmethyl)-1,4,7-triazacyclononane, whose identity was
35 established by proton NMR.

Those skilled in the art will recognize that this procedure can be used to place other alkyl or aryl halides on the methylene carbon as well, using the appropriate alkyl or aryl halide.

C. Hydrolysis of the Perethyl Esters to the Phosphonic Acids

In this procedure, both perethyl esters of part A above were converted to the corresponding phosphonic acids.

The perethyl esters were separately dissolved in concentrated hydrochloric acid and heated at 80°C for six to eight hours. The resulting solutions were evaporated to dryness, and the pure acid forms were obtained following crystallization from water or water/ethanol. Their identity as the acids was confirmed by proton NMR and elemental analysis.

To further confirm the identity of the products, independent syntheses were performed employing the method described by Polykarpov, Yu.M., et al., "N,N',N''-Tris(phosphonomethyl)-1,4,7-triazacyclononane -- a specific complexing agent for magnesium ion," *Izv. Akad. Nauk SSSR. Ser. Khim.*, 1982, (7), 1669-70. The products obtained were found to be identical by NMR to those obtained by the synthesis described above.

EXAMPLE 2

SYNTHESIS OF DIHYDROXYPHOSPHORYLETHYL SPECIES

This example illustrates the preparation of certain dihydroxyphosphorylethyl analogs of the compounds prepared in Example 1. Species based on both 1,4,7-triazacyclononane and 1,4,7,10-tetraazacyclododecane are again illustrated in parallel fashion.

A. Synthesis of Per-N-Substituted Dihydroxyphosphorylethyl Phosphonates

Diethyl 2-bromoethylphosphonate, prepared by procedures described in the literature, was reacted separately with the hydrobromide salts of 1,4,7-triazacyclononane and 1,4,7,10-tetraazacyclododecane in water in the presence of excess K_2CO_3 at $80^\circ C$ for 4-5 hours. The water was then removed by evaporation and chloroform was added to the solids to remove the product from the inorganic salts. The products were purified by chromatography employing neutral alumina and an elution solvent of 10% methanol in chloroform. The perethyl ester groups were removed by hydrolysis using HCl as described in part C of Example 1 above. The pure products were obtained by crystallization from 10% ethanol in water, and their identity was established as N,N',N''-tris(dihydroxyphosphorylethyl)-1,4,7-triazacyclononane and N,N',N'',N'''-tetrakis(dihydroxyphosphorylethyl)-1,4,7,10-tetraazacyclododecane, respectively, by proton NMR and elemental analysis after accounting for waters of hydration.

Those skilled in the art will recognize that this procedure can be employed in an analogous manner to synthesize similar compounds having alkyl or aryl substitutions on the ethylene carbon atoms by employing correspondingly substituted diethyl 2-bromoethylphosphonate.

B. Synthesis of N,N',N''-tris(dihydroxyphosphorylhydroxyethyl)-1,4,7-triazacyclononane

Diethyl epoxyethyl phosphonate, prepared employing known procedures, was combined with a solution of 1,4,7-triazacyclononane in methanol at room temperature, using 3.3 equivalents of the diethyl epoxyethyl phosphonate. The solution was stirred for six hours at $40-50^\circ C$. The methanol was evaporated and the residue was dissolved in water. The excess epoxide was then extracted with diethyl ether and the water layer was evaporated. The residue was purified by chromatography using neutral alumina by first eluting the

column with chloroform followed by 10% methanol in chloroform. The perethyl ester groups were removed by hydrolysis in HCl as described above. The pure product was obtained by crystallization from 10% ethanol in water. The identity of the product was established as that of N,N',N''-tris(dihydroxyphosphorylhydroxyethyl)-1,4,7-triazacyclononane by proton NMR and elemental analysis after accounting for three molecules of water of hydration.

Those skilled in the art will recognize that the tris-dihydroxyphosphorylaminoethyl analog is similarly prepared by the same procedure, using diethyl ethylenimino phosphonate in place of the diethyl epoxyethyl phosphonate, and that similar compounds bearing alkyl or aryl substitutions on the ethylene carbon atoms are prepared analogously by employing correspondingly substituted forms of diethyl epoxyethyl phosphonate or diethyl ethylenimino phosphonate. The same procedure can likewise be used to synthesize hydroxyphosphorylhydroxyethyl and hydroxyphosphorylaminoethyl derivatives of other polyamines.

EXAMPLE 3

PREPARATION OF METAL CATION COMPLEXES

A. Fe(III) and Cr(III) Complexes of N,N',N''-Tris(dihydroxyphosphorylmethyl)-1,4,7-triazacyclononane

In separate syntheses, N,N',N''-tris(dihydroxyphosphorylmethyl)-1,4,7-triazacyclononane in water was combined with an equivalent of a water-soluble salt of the appropriate metal cation to be included in the complex (i.e., $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ or $\text{CrCl}_3 \cdot 6\text{H}_2\text{O}$). The mixtures were heated in a reflux apparatus at 100°C while base was slowly added in an amount equal to "n" equivalents, where "n" equals the charge of the metal cation. The Fe(III) complex crystallized from the aqueous solution, permitting recovery in high yield. The Cr(III) complex crystallized upon cooling.

addition of ethanol. The crystallized complex in each case was added to fresh water and sufficient base was added to yield a final pH of >10.0 . In the case of the Fe(III) complex, the resulting solution was heated to 100°C while in the case of the Cr(III) complex, the resulting solution was heated to 140°C (under pressure). Heating was continued in each case until a single chromatographic species was obtained. The solutions were then cooled and filtered to remove solids, and acid was added to the filtrate to crystallize or precipitate the complex as before. Additional crystallizations of the complex were performed from water or water/ethanol. The purity of each complex was established by thin layer chromatography (TLC). The identity of each product was established by elemental analysis.

B. Fe(III) Complexes of N,N',N'' -Tris(dihydroxyphosphorylethyl)-1,4,7-triazacyclononane and N,N',N'' -Tris(dihydroxyphosphorylhydroxyethyl)-1,4,7-triazacyclononane.

These complexes were prepared following modifications of the procedure of part A above. When solutions of the Fe(III) complex of N,N',N'' -tris(dihydroxyphosphorylethyl)-1,4,7-triazacyclononane were acidified, additional products were noted on chromatographic analysis. Consequently, the final recrystallization step requiring acidification was eliminated, and the neutral form of the complex was not isolated as a solid. The tri-sodium salt of the product was purified in an ion exchange column, and the inorganic salts were removed by use of an LH20 column. In the case of the Fe(III) complex of N,N',N'' -tris(dihydroxyphosphorylhydroxyethyl)-1,4,7-triazacyclononane, a single chromatographically distinct product was not obtained using this procedure.

C. Mn(II) and Mn(III) Complexes of N,N',N''-Tris-(
(dihydroxyphosphorylmethyl)-1,4,7-triazacyclononane

5 The procedure for these complexes was modified due to the ease with which redox reactions occurred. The tetra-sodium salt of the Mn(II) complex was formed directly by adding six equivalents of NaOH to a 1:1 mixture of ligand and MnCl₂ in water, followed by crystallization of the salt of the complex by addition of ethanol and cooling. To prepare the tri-sodium salt of the Mn(III) complex, a stoichiometric quantity of persulfate ion was added to the tetra-sodium salt of the Mn(II) complex, and the reaction allowed to stand at room temperature until all of the Mn(II) had oxidized to Mn(III). The product was purified by passage through an ion exchange column, and inorganic salts were removed by passage through an LH-20 column. Both products were characterized as single, chromatographically distinct products on TLC.

20 D. Gd(III) Complexes of N,N',N''-Tris(dihydroxyphosphorylmethyl)-1,4,7-triazacyclononane,
N,N',N'',N'''-Tetrakis(dihydroxyphosphorylmethyl)-1,4,7,10-tetraazacyclododecane and N,N',N'',N'''-Tetrakis(dihydroxyphosphorylethyl)-1,4,7,10-tetraazacyclododecane

25 The tri-sodium salt of the Gd(III) complex of N,N',N''-tris(dihydroxyphosphorylmethyl)-1,4,7-triazacyclononane was made directly from GdCl₃·6H₂O and the acid form of the ligand by adding equivalent amounts of each to water and heating at 100°C. When the solution was clear, six equivalents of NaOH were added slowly, and the solution was heated for an additional five days. After centrifugation to remove the small amount of residual solids, the solution was dried to give a solid which was

characterized as a single, chromatographically distinct product on TLC.

The neutral form of the Gd(III) complex of N,N',N'',N'''-tetrakis(dihydroxyphosphorylmethyl)-1,4,7,10-tetraazacyclododecane was made from $\text{GdCl}_3 \cdot 6\text{H}_2\text{O}$ and the acid form of the ligand by adding equivalent amounts of each to water, followed by slow addition of three equivalents of NaOH. Heating at 90°C resulted in formation of a gelatinous precipitate. Heating was continued until no further precipitate formed, and the reaction mixture was allowed to cool to room temperature. The precipitate which formed as a result was isolated by centrifugation and washed with water. The washed precipitate was added to water, the pH was raised above 11 by addition of NaOH, and the resulting clear solution was heated overnight at 100°C. The solution was acidified to pH<3.0 with concentrated HCl, and was concentrated and cooled, yielding solids which were separated by centrifugation.

The pentameglumine salt of the Gd(III) complex of N,N',N'',N'''-tetrakis(dihydroxyphosphorylethyl)-1,4,7,10-tetraazacyclododecane was made directly from Gd_2O_3 and the acid form of the ligand by adding 0.5 molecular equivalents of the former and 1.0 molecular equivalents of the latter to water and heating at 90°C until a clear solution was obtained. After filtration, five equivalents of meglumine were added to the filtrate, and the reaction was heated at 100°C for 20 hours. After cooling, the reaction mixture was brought to dryness to obtain the solid product.

EXAMPLE 4

PREPARATION OF PHYSIOLOGICAL SALTS

A. Sodium and Meglumine Salts of Fe(III) Complex with
5 N,N',N''-Tris(dihydroxyphosphorylmethyl)-1,4,7-
triazacyclononane.

The solid form of the Fe(III) complex of N,N',N''-
tris(dihydroxyphosphorylmethyl)-1,4,7-triazacyclononane, the
10 complex whose preparation is described in Example 3, part A
above, was dissolved in water at room temperature. Sodium
hydroxide or meglumine were added to separate solutions of
the neutral Fe(III) complex of N,N',N''-tris(dihydroxy-
phosphorylmethyl)-1,4,7-triazacyclononane until the
15 solutions maintained a pH of 7.0 to 7.4. The solutions were
then lyophilized to obtain the solid physiological salts of
the complex. These solids, when reconstituted with a
suitable aqueous solvent prior to use, are suitable for in
vivo administration. In each case, for the sodium salts of
20 the complexes, potentiometric titration demonstrated that
the principal form of the complexes at pH 7.0 to 7.4 was the
trianion of the complex, and thus that the principal salt
forms at this pH were the trisodium and the trimeglumine
salt.

25 B. Meglumine Salt of Cr(III) Complex with N,N',N''-
Tris(dihydroxyphosphorylmethyl)-1,4,7-triaza-
cyclononane

30 The solid form of the Cr(III) complex of N,N',N''-
tris(dihydroxyphosphorylmethyl)-1,4,7-triazacyclononane, the
complex whose preparation is described in Example 3, part A
above, was treated with three equivalents of meglumine in a
fashion comparable to that described in Example 4, Part A
35 above. Potentiometric titration of the sodium salt of this
complex demonstrated that the principal form of the
resulting salt at pH 7.0 to 7.4 was the trianion.

Those skilled in the art will recognize that other salts of the subject complexes can be obtained employing similar procedures.

5

EXAMPLE 5

PRODUCT EVALUATION

10 In the following studies, the test species are referred to as follows:

1. <i>Salmonella typhimurium</i>	2. <i>Salmonella typhimurium</i>
3. <i>Salmonella typhimurium</i>	4. <i>Salmonella typhimurium</i>
5. <i>Salmonella typhimurium</i>	6. <i>Salmonella typhimurium</i>
7. <i>Salmonella typhimurium</i>	8. <i>Salmonella typhimurium</i>
9. <i>Salmonella typhimurium</i>	10. <i>Salmonella typhimurium</i>
11. <i>Salmonella typhimurium</i>	12. <i>Salmonella typhimurium</i>
13. <i>Salmonella typhimurium</i>	14. <i>Salmonella typhimurium</i>
15. <i>Salmonella typhimurium</i>	16. <i>Salmonella typhimurium</i>
17. <i>Salmonella typhimurium</i>	18. <i>Salmonella typhimurium</i>
19. <i>Salmonella typhimurium</i>	20. <i>Salmonella typhimurium</i>
21. <i>Salmonella typhimurium</i>	22. <i>Salmonella typhimurium</i>
23. <i>Salmonella typhimurium</i>	24. <i>Salmonella typhimurium</i>
25. <i>Salmonella typhimurium</i>	26. <i>Salmonella typhimurium</i>
27. <i>Salmonella typhimurium</i>	28. <i>Salmonella typhimurium</i>
29. <i>Salmonella typhimurium</i>	30. <i>Salmonella typhimurium</i>
31. <i>Salmonella typhimurium</i>	32. <i>Salmonella typhimurium</i>
33. <i>Salmonella typhimurium</i>	34. <i>Salmonella typhimurium</i>
35. <i>Salmonella typhimurium</i>	36. <i>Salmonella typhimurium</i>
37. <i>Salmonella typhimurium</i>	38. <i>Salmonella typhimurium</i>
39. <i>Salmonella typhimurium</i>	40. <i>Salmonella typhimurium</i>
41. <i>Salmonella typhimurium</i>	42. <i>Salmonella typhimurium</i>
43. <i>Salmonella typhimurium</i>	44. <i>Salmonella typhimurium</i>
45. <i>Salmonella typhimurium</i>	46. <i>Salmonella typhimurium</i>
47. <i>Salmonella typhimurium</i>	48. <i>Salmonella typhimurium</i>
49. <i>Salmonella typhimurium</i>	50. <i>Salmonella typhimurium</i>
51. <i>Salmonella typhimurium</i>	52. <i>Salmonella typhimurium</i>
53. <i>Salmonella typhimurium</i>	54. <i>Salmonella typhimurium</i>
55. <i>Salmonella typhimurium</i>	56. <i>Salmonella typhimurium</i>
57. <i>Salmonella typhimurium</i>	58. <i>Salmonella typhimurium</i>
59. <i>Salmonella typhimurium</i>	60. <i>Salmonella typhimurium</i>
61. <i>Salmonella typhimurium</i>	62. <i>Salmonella typhimurium</i>
63. <i>Salmonella typhimurium</i>	64. <i>Salmonella typhimurium</i>
65. <i>Salmonella typhimurium</i>	66. <i>Salmonella typhimurium</i>
67. <i>Salmonella typhimurium</i>	68. <i>Salmonella typhimurium</i>
69. <i>Salmonella typhimurium</i>	70. <i>Salmonella typhimurium</i>
71. <i>Salmonella typhimurium</i>	72. <i>Salmonella typhimurium</i>
73. <i>Salmonella typhimurium</i>	74. <i>Salmonella typhimurium</i>
75. <i>Salmonella typhimurium</i>	76. <i>Salmonella typhimurium</i>
77. <i>Salmonella typhimurium</i>	78. <i>Salmonella typhimurium</i>
79. <i>Salmonella typhimurium</i>	80. <i>Salmonella typhimurium</i>
81. <i>Salmonella typhimurium</i>	82. <i>Salmonella typhimurium</i>
83. <i>Salmonella typhimurium</i>	84. <i>Salmonella typhimurium</i>
85. <i>Salmonella typhimurium</i>	86. <i>Salmonella typhimurium</i>
87. <i>Salmonella typhimurium</i>	88. <i>Salmonella typhimurium</i>
89. <i>Salmonella typhimurium</i>	90. <i>Salmonella typhimurium</i>
91. <i>Salmonella typhimurium</i>	92. <i>Salmonella typhimurium</i>
93. <i>Salmonella typhimurium</i>	94. <i>Salmonella typhimurium</i>
95. <i>Salmonella typhimurium</i>	96. <i>Salmonella typhimurium</i>
97. <i>Salmonella typhimurium</i>	98. <i>Salmonella typhimurium</i>
99. <i>Salmonella typhimurium</i>	100. <i>Salmonella typhimurium</i>

TABLE 1
Test Species

Ref.	Ligand	Para-magnetic Cation	Physio- logically Compatible Cation
A	N,N',N''-tris(dihydroxy-phosphorylmethyl)-1,4,7-triazacyclononane	Fe(III)	trisodium
B	N,N',N''-tris(dihydroxy-phosphorylmethyl)-1,4,7-triazacyclononane	Fe(III)	tri-meglumine
C	N,N',N''-tris(dihydroxy-phosphorylethyl)-1,4,7-triazacyclononane	Fe(III)	tri-sodium
D	N,N',N''-tris(dihydroxy-phosphorylmethyl)-1,4,7-triazacyclononane	Cr(III)	tri-meglumine
E	N,N',N''-tris(dihydroxy-phosphorylmethyl)-1,4,7-triazacyclononane	Mn(II)	tetra-sodium
F	N,N',N''-tris(dihydroxy-phosphorylmethyl)-1,4,7-triazacyclononane	Mn(III)	tri-sodium
G	N,N',N''-tris(dihydroxy-phosphorylmethyl)-1,4,7-triazacyclononane	Gd(III)	tri-sodium
H	N,N',N'',N'''-tetrakis-(dihydroxyphosphorylethyl)-1,4,7,10-tetraazacyclododecane	Gd(III)	penta-meglumine

A. Water Solubility

All of the test species listed above were dissolved in water, demonstrating solubility at concentrations sufficient to be useful as pharmaceutical agents. In particular, test species A and B proved soluble in water at concentrations exceeding 50% (weight/volume).

B. Stability

TLC was performed on test species A and D, both before and after heating at 100°C for two hours. A single, chromatographically distinct spot which did not vary as a result of the heating was observed in both cases.

C. Toxicity

Physiological salts of the various ligand/metal cation complexes described herein were administered intravenously to mice. The mice were observed for two weeks following such administration, and the results are listed in Table 2 below. In this data, the administered dose is expressed as mM of complex per kg whole body weight (mM/kg), and the administration rate is expressed as mM of complex administered per second or per minute per kg whole body weight (mM/sec/kg or mM/min/kg). The mice were considered to have "survived" administration of each agent if they were alive at the end of the two-week period.

TABLE 2
Toxicity Test Results

Test	Species	Dose	Rate	Survived?
(1)	A	11.8mM/kg	0.4mM/sec/kg	yes
(2)	B	9.8mM/kg	0.7mM/min/kg	yes
(3)	C	10.0mM/kg	2.0mM/min/kg	yes
(4)	D	2.9mM/kg	0.5mM/sec/kg	yes
(5)	E	2.9mM/kg	1.1mM/min/kg	yes
(6)	F	8.0mM/kg	2.6mM/min/kg	yes
(7)	G	3.1mM/kg	0.8mM/min/kg	yes

When complexes of Fe(III) with N,N',N''-tris-(dihydroxyphosphorylmethyl)-1,4,7-triazacyclononane (Test

Species A) were prepared employing alternate complexation procedures and which contained multiple forms of the complex, as detected by TLC; the toxicity observed was substantially greater than that observed with preparations of this complex prepared employing the procedure described herein and which showed a single, chromatographically distinct product on TLC.

D. In Vivo Distribution And Whole Body Clearance

Studies
Radioisotopically labeled analogs of test species selected from those listed above were synthesized, using radioisotopes of Fe(III) (iron-59), Cr(III) (chromium-51) and Gd(III) (gadolinium-153), and employing the general synthesis procedures described above. The resulting complexes were subjected to radiochromatography to insure acceptable radiopurity and identity with the parent complexes. They were then administered intravenously to mice in order to measure in vivo distribution and whole body clearance.

The location of the test species in the mice's bodies and the rate at which the test species were cleared from the mice's bodies after administration were determined by radioassay of tissues and whole body counting, both performed by conventional gamma ray counting techniques. Concentration of activity in tissues was determined as activity per gram of tissue, and whole body counts were expressed as the percentage of whole body activity at a given time with respect to whole body activity immediately after injection. The results were as follows:

1. Radioisotopes of Test Species A and B.

In vivo distribution. Within 2 minutes

following administration, evidence of

concentration of radioactivity in bone and kidneys was seen. By measurements taken

one hour after administration, the ratio of the concentration of the test species in bone to that in whole blood was generally over 25:1, while the ratio of their concentration in kidney to that in blood was generally over 10:1. Even after 24 hours, when less than 5% of the administered dose remained in the body (see below), a high ratio of concentration of the test species in bone and kidney to that in blood was maintained. In mice who had a tibia broken two to four weeks prior to the study, the tibia which had suffered the fracture showed significantly greater accumulation of activity than that which was measured in the contralateral normal tibia. All mice showed very low activity in the brain at all times following their administration.

Whole Body Clearance. Within 24 hours of administration, over 95% of both test species had been cleared from the body, almost exclusively through the urine.

2. Radioisotope of Test Species C.

In vivo distribution. One hour after administration, the measured bone-to-blood concentration ratio was greater than 4.5:1, and the measured kidney-to-blood ratio was greater than 5.5:1. A very low concentration of this agent in brain was noted within this first hour.

Whole Body Clearance. Within 24 hours of administration, over 95% of the test species had been cleared from the body.

3. Radioisotope of Test Species D.

In vivo distribution. One hour after

administration, the measured bone-to-blood concentration ratio was greater than 6:1, and the measured kidney-to-blood ratio was greater than 10:1. A very low concentration of agent in brain was noted within this first hour.

Whole Body Clearance. Within 24 hours of

administration, over 95% of the test species had been cleared from the body, almost exclusively through the urine.

E. Relaxivity Measurements.

Measurements of proton longitudinal relaxivity ($1/T_1$) were performed on some of the test species listed above, and compared with those obtained using the following complexes outside the scope of this invention: (i) Gd(III) with diethylenetriamine pentaacetic acid (DTPA), and (ii) Fe(III) with N,N'-ethylenbis[(2-hydroxyphenyl)-glycinate] (EHPG). All measurements were obtained using a Bruckner PC/20 Minispec device operating at 20 MHz. All samples were dissolved in 0.1 M phosphate buffer at pH 7.2.

The proton longitudinal relaxivity of Test Species A was two to three times greater than that obtained for the Fe(III) complex of EHPG and between 40 and 45% of that obtained for the Gd(III) complex of DTPA. Similar results were obtained for Test Species C.

F. Relative Equilibrium Constant Measurements

The highly colored Fe(III) complex of EHPG (used for comparison in part E of this example) was dissolved in 0.25 M phosphate buffer at pH 7.2, and an equimolar quantity of N,N',N''-tris(dihydroxyphosphorylmethyl)-1,4,7-triazacyclononane was added. The solution was heated

overnight at 100°C. The resulting solution was devoid of the purple Fe(III) EHPG complex color, and TLC showed only the presence of the Fe(III) N,N',N''-tris(dihydroxyphosphorylmethyl)-1,4,7-triazacyclononane complex.

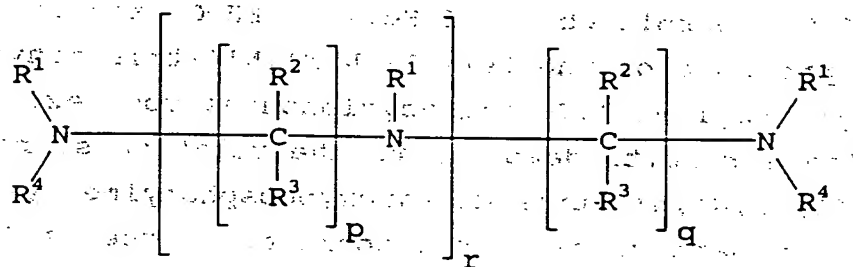
5 The reverse experiment was also performed. The Fe(III) complex of N,N',N''-tris(dihydroxyphosphorylmethyl)-1,4,7-triazacyclononane was thus dissolved in 0.25M phosphate buffer at pH 7.2, and an equimolar quantity of EHPG was added. The solution was heated overnight at 100°C.
10 As in the first experiment, the resulting solution was devoid of the purple color of Fe(III) EHPG, and TLC showed only the presence of the Fe(III) N,N',N''-tris(dihydroxyphosphorylmethyl)-1,4,7-triazacyclononane complex.

15 These results demonstrate the relative stability of the Fe(III) N,N',N''-tris(dihydroxyphosphorylmethyl)-1,4,7-triazacyclononane complex in comparison to the Fe(III) EHPG complex.

20 The foregoing is offered primarily for purposes of illustration. It will be readily apparent to those skilled in the art that further variations, substitutions and modifications in the substances and procedures involved in the invention beyond those specifically disclosed herein may be made without departing from the spirit and scope of the
25 invention.

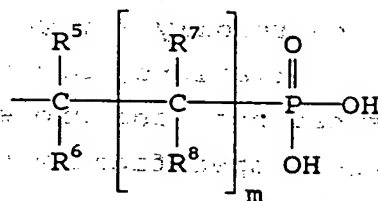
WHAT IS CLAIMED IS:

1. A method of preferentially enhancing magnetic resonance image contrast in bone tissue and other tissue of a patient bearing recognition features in common with bone tissue, said method comprising administering to said patient an effective amount of a pharmaceutical agent comprising a physiologically compatible salt of a chelate of a paramagnetic metal cation and a compound having the formula



in which:

the R¹ moieties are each independently



in which R⁵, R⁶ and R⁷ are independently selected from the group consisting of H and alkyl and aryl groups which do not interfere with complexation; R⁸ is selected from the group consisting of H, OH, NH₂, and alkyl and aryl groups which do not interfere with complexation; and m is zero or 1;

the R² moieties are each independently selected from the group consisting of H and alkyl and aryl groups which do not interfere with complexation;

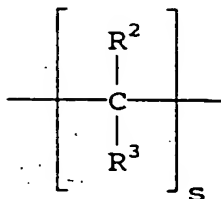
the R³ moieties are each independently selected from the group consisting of H and alkyl and aryl groups which do not interfere with complexation;

p is 2 or 3;

q is 2 or 3;

r is 0 to 3; and

the R⁴ moieties are either independently selected from the the definition of the R¹ moieties or together form a single divalent group having the formula



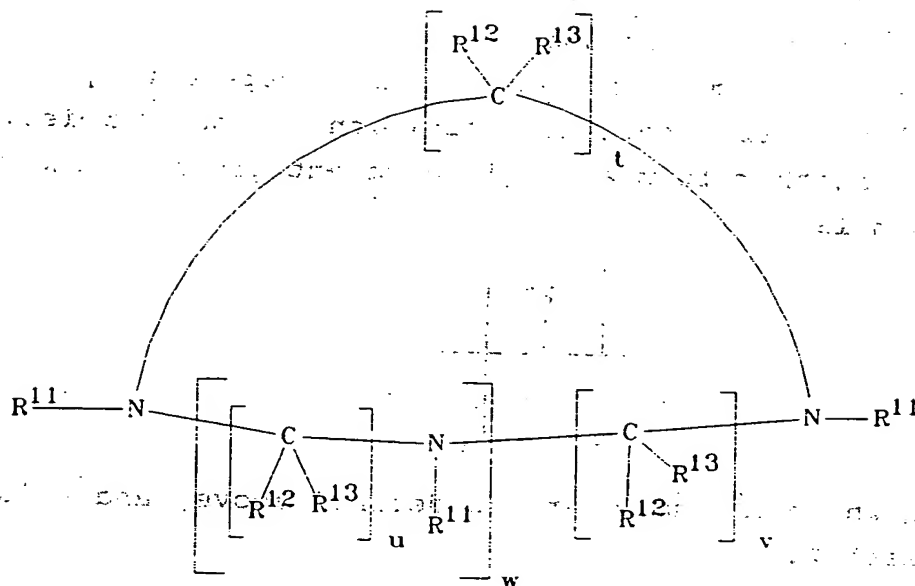
in which R² and R³ are as defined above, and s is at least 2.

2. A method in accordance with claim 1 in which r is 0 to 2.

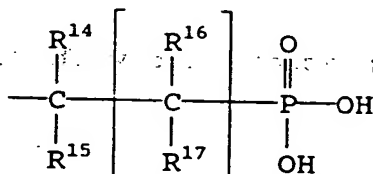
3. A method in accordance with claim 1 in which r is 0 or 1.

4. A method in accordance with claim 1 in which s is 2 or 3.

5. A method of preferentially enhancing magnetic resonance image contrast in bone tissue and other tissue of a patient bearing recognition features in common with bone tissue, said method comprising administering to said patient an effective amount of a pharmaceutical agent comprising a physiologically compatible salt of a chelate of a paramagnetic metal cation and a compound having the formula



in which: the R^{11} moieties are each independently



in which R^{14} , R^{15} and R^{16} are independently selected from the group consisting of H and alkyl and aryl groups which do not interfere with complexation; R^{17} is selected from the group consisting of H, OH, NH_2 , and alkyl and aryl groups which do not interfere with complexation; and n is zero or 1;

the R^{12} moieties are each independently selected from the group consisting of H and alkyl and aryl groups which do not interfere with complexation;

the R^{13} moieties are each independently selected from the group consisting of H and alkyl and aryl groups which do not interfere with complexation;

t is 2 or 3;

u is 2 or 3;

v is 2 or 3; and

w is at least 1.

6. A method in accordance with claim 5 in which R^{14} , R^{15} and R^{16} are independently selected from the group consisting of H, C_1-C_8 alkyl, phenyl and benzyl; and R^{17} is
5 selected from the group consisting of H, OH, NH_2 , C_1-C_8 alkyl, phenyl and benzyl.

7. A method in accordance with claim 5 in which R^{14} , R^{15} and R^{16} are independently selected from the group
10 consisting of H, C_1-C_4 alkyl and benzyl; and R^{17} is selected from the group consisting of H, OH, NH_2 , C_1-C_4 alkyl and benzyl.

8. A method in accordance with claim 5 in which
15 R^{14} , R^{15} and R^{16} are each H; R^{17} is selected from the group consisting of H, OH, NH_2 , C_1-C_8 alkyl, phenyl and benzyl; and n is 1.

9. A method in accordance with claim 5 in which
20 R^{14} , R^{15} and R^{16} are each H; R^{17} is selected from the group consisting of H, OH, NH_2 , C_1-C_4 alkyl and benzyl; and n is 1.

10. A method in accordance with claim 5 in which
 R^{14} , R^{15} , R^{16} and R^{17} are each H; and n is 1.

11. A method in accordance with claim 5 in which R^{14}
and R^{15} are independently selected from the group consisting
of H, C_1-C_8 alkyl, phenyl and benzyl; and n is zero.

12. A method in accordance with claim 5 in which R^{14}
30 and R^{15} are independently selected from the group consisting
of H, C_1-C_4 alkyl and benzyl; and n is zero.

13. A method in accordance with claim 5 in which R^{14}
35 and R^{15} are each H; and n is zero.

14. A method in accordance with claim 5 in which the R^{12} and R^{13} moieties are each independently selected from the group consisting of H, C_1-C_8 alkyl, phenyl and benzyl.

5 15. A method in accordance with claim 5 in which the R^{12} and R^{13} moieties are each independently selected from the group consisting of H, C_1-C_4 alkyl and benzyl.

10 16. A method in accordance with claim 5 in which the R^{12} moieties are each H; and the R^{13} moieties are each independently selected from the group consisting of H, C_1-C_8 alkyl, phenyl and benzyl.

15 17. A method in accordance with claim 5 in which the R^{12} moieties are each H; and the R^{13} moieties are each independently selected from the group consisting of H, C_1-C_4 alkyl and benzyl.

20 18. A method in accordance with claim 5 in which the R^{12} moieties are each H; and the R^{13} moieties are each independently selected from the group consisting of H and C_1-C_4 alkyl.

25 19. A method in accordance with claim 5 in which the R^{12} moieties are each H; and the R^{13} moieties are each independently selected from the group consisting of H and methyl.

30 20. A method in accordance with claim 5 in which the R^{12} moieties are each H; and the R^{13} moieties are each H.

21. A method in accordance with claim 5 in which t, u and v are each 2.

35 22. A method in accordance with claim 5 in which w is 1 to 4.

23. A method in accordance with claim 5 in which w is 1 to 3.

24. A method in accordance with claim 5 in which w is 1.

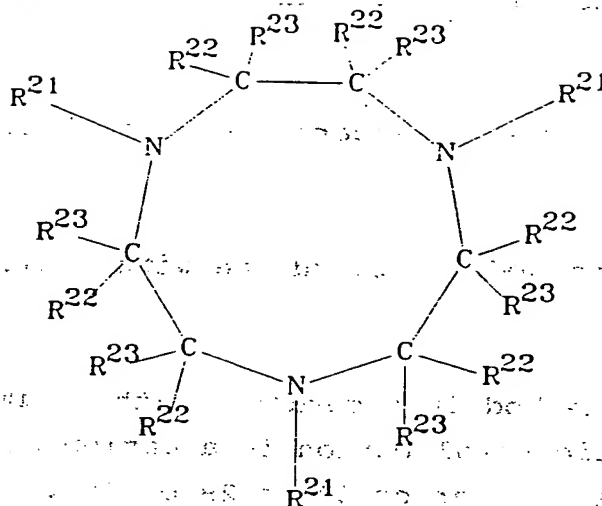
25. A method in accordance with claim 5 in which w is 2.

26. A method in accordance with claim 5 in which said paramagnetic metal cation is a cation of an element having an atomic number of 22 to 29 or 58 to 70.

27. A method in accordance with claim 5 in which said paramagnetic metal cation is a cation of an element selected from the group consisting of chromium, manganese, iron and gadolinium.

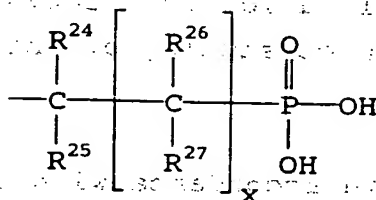
28. A method in accordance with claim 5 in which said physiological compatible salt is comprised of said chelate in combination with at least one cation selected from the group consisting of sodium and N-methylglucamine.

29. A method of preferentially enhancing magnetic resonance image contrast in bone tissue and other tissue of a patient bearing recognition features in common with bone tissue, said method comprising administering to said patient an effective amount of a pharmaceutical agent comprising a physiologically compatible salt of a chelate of a paramagnetic metal cation and a compound having the formula



in which:

the R^{21} moieties are each independently



in which R^{24} , R^{25} and R^{26} are independently selected from the group consisting of H and alkyl and aryl groups which do not interfere with complexation; R^{27} is selected from the group consisting of H, OH, NH_2 , and alkyl and aryl groups which do not interfere with complexation; and x is zero or 1;

the R^{22} moieties are each independently selected from the group consisting of H and alkyl and aryl groups which do not interfere with complexation; and the R^{23} moieties are each independently selected from the group consisting of H and alkyl and aryl groups which do not interfere with complexation.

30. A method in accordance with claim 29 in which R^{24} , R^{25} and R^{26} are independently selected from the group consisting of H, C_1 - C_4 alkyl and benzyl; and R^{27} is selected from the group consisting of H, OH, NH_2 , C_1 - C_4 alkyl and benzyl.

31. A method in accordance with claim 29 in which R^{24} , R^{25} and R^{26} are each H; R^{27} is selected from the group consisting of H, OH, NH_2 , C_1-C_4 alkyl and benzyl; and x is 1.

32. A method in accordance with claim 29 in which R^{24} , R^{25} , R^{26} and R^{27} are each H; and x is 1.

33. A method in accordance with claim 29 in which R^{24} and R^{25} are independently selected from the group consisting of H, C_1-C_8 alkyl, phenyl and benzyl; and x is zero.

34. A method in accordance with claim 29 in which R^{24} and R^{25} are independently selected from the group consisting of H, C_1-C_4 alkyl and benzyl; and x is zero.

35. A method in accordance with claim 29 in which R^{24} and R^{25} are each H; and x is zero.

36. A method in accordance with claim 29 in which the R^{22} and R^{23} moieties are each independently selected from the group consisting of H, C_1-C_4 alkyl and benzyl.

37. A method in accordance with claim 29 in which the R^{22} moieties are each H; and the R^{23} moieties are each independently selected from the group consisting of H and C_1-C_4 alkyl.

38. A method in accordance with claim 29 in which the R^{22} moieties are each H; and the R^{23} moieties are each independently selected from the group consisting of H and methyl.

39. A method in accordance with claim 29 in which the R^{22} moieties are each H; and the R^{23} moieties are each H.

40. A method in accordance with claim 29 in which R^{24} and R^{25} are each H; x is zero; the R^{22} moieties are each H; and the R^{23} moieties are each H.

5 41. A method in accordance with claim 29 in which R^{24} , R^{25} , R^{26} and R^{27} are each H; x is 1; the R^{22} moieties are each H; and the R^{23} moieties are each H.

10 42. A method in accordance with claim 29 in which R^{24} , R^{25} and R^{26} are each H; R^{27} is OH; x is 1; the R^{22} moieties are each H; and the R^{23} moieties are each H.

15 43. A method in accordance with claim 29 in which R^{24} , R^{25} and R^{26} are each H; R^{27} is NH_2 ; x is 1; the R^{22} moieties are each H; and the R^{23} moieties are each H.

20 44. A method in accordance with claim 29 in which said paramagnetic metal cation is a cation of an element having an atomic number of 22 to 29 or 58 to 70.

25 45. A method in accordance with claim 29 in which said paramagnetic metal cation is a cation of an element selected from the group consisting of chromium, manganese, iron and gadolinium.

30 46. A method in accordance with claim 29 in which said physiological compatible salt is comprised of said chelate in combination with at least one cation selected from the group consisting of sodium and N-methylglucamine.

35 47. A method in accordance with claim 29 in which said physiologically compatible salt is the combination of three equivalents of a physiologically compatible cation with the trianionic complex of Fe(III) and N,N',N'''-tris(di-hydroxyphosphorylmethyl)-1,4,7-triazacyclononane at a pH of about 6.8 to about 7.4.

48. A method in accordance with claim 29 in which said physiologically compatible salt is the trisodium salt of the Fe(III) complex of N,N',N''-tris(dihydroxyphosphorylmethyl)-1,4,7-triazacyclononane.

49. A method in accordance with claim 29 in which said physiologically compatible salt is the trimeglumine salt of the Fe(III) complex of N,N',N''-tris(dihydroxyphosphorylmethyl)-1,4,7-triazacyclononane.

50. A method in accordance with claim 29 in which said physiologically compatible salt is the trisodium salt of the Fe(III) complex of N,N',N''-tris(dihydroxyphosphoryl-ethyl)-1,4,7-triazacyclononane.

51. A method in accordance with claim 29 in which said physiologically compatible salt is the trimeglumine salt of the Cr(III) complex of N,N',N''-tris(dihydroxyphosphorylmethyl)-1,4,7-triazacyclononane.

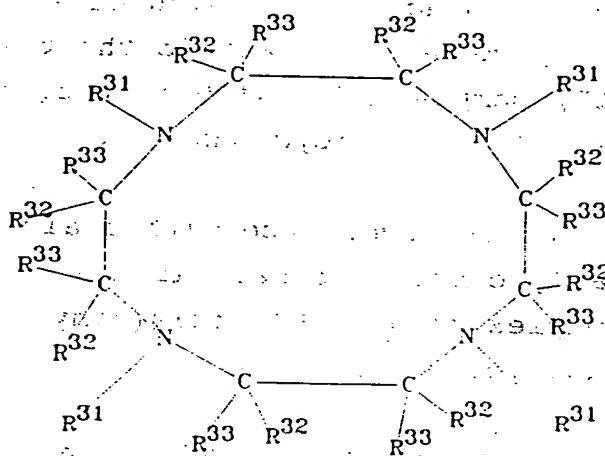
52. A method in accordance with claim 29 in which said physiologically compatible salt is the tetrasodium salt of the Mn(II) complex of N,N',N''-tris(dihydroxyphosphorylmethyl)-1,4,7-triazacyclononane.

53. A method in accordance with claim 29 in which said physiologically compatible salt is the trisodium salt of the Mn(III) complex of N,N',N''-tris(dihydroxyphosphorylmethyl)-1,4,7-triazacyclononane.

54. A method in accordance with claim 29 in which said physiologically compatible salt is the trisodium salt of the Gd(III) complex of N,N',N''-tris(dihydroxyphosphorylmethyl)-1,4,7-triazacyclononane.

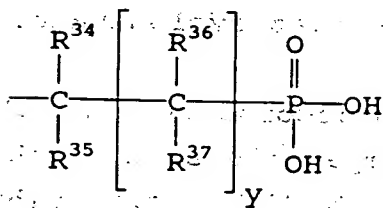
55. A method of preferentially enhancing magnetic resonance image contrast in bone tissue and other tissue of

a patient bearing recognition features in common with bone tissue, said method comprising administering to said patient an effective amount of a pharmaceutical agent comprising a physiologically compatible salt of a chelate of a
 5 paramagnetic metal cation and a compound having the formula



in which:

the R^{31} moieties are each independently



in which R^{34} , R^{35} and R^{36} are independently selected

from the group consisting of H and alkyl and aryl

groups which do not interfere with complexation; R^{37}

is selected from the group consisting of H, OH, NH_2 ,

and alkyl and aryl groups which do not interfere

with complexation; and y is zero or 1;

the R^{32} moieties are each independently selected

from the group consisting of H and alkyl and aryl

groups which do not interfere with complexation; and

the R^{33} moieties are each independently selected

from the group consisting of H and alkyl and aryl

groups which do not interfere with complexation.

56. A method in accordance with claim 55 in which R^{34} , R^{35} and R^{36} are independently selected from the group consisting of H, C_1-C_4 alkyl and benzyl; and R^{37} is selected from the group consisting of H, OH, NH_2 , C_1-C_4 alkyl and benzyl.

57. A method in accordance with claim 55 in which R^{34} , R^{35} and R^{36} are each H; R^{37} is selected from the group consisting of H, OH, NH_2 , C_1-C_4 alkyl and benzyl; and y is 1.

58. A method in accordance with claim 55 in which R^{34} , R^{35} , R^{36} and R^{37} are each H; and y is 1.

59. A method in accordance with claim 55 in which R^{34} and R^{35} are independently selected from the group consisting of H, C_1-C_4 alkyl and benzyl; and y is zero.

60. A method in accordance with claim 55 in which R^{34} and R^{35} are each H; and y is zero.

61. A method in accordance with claim 55 in which the R^{32} and R^{33} moieties are each independently selected from the group consisting of H, C_1-C_4 alkyl and benzyl.

62. A method in accordance with claim 55 in which the R^{32} moieties are each H; and the R^{33} moieties are each independently selected from the group consisting of H, C_1-C_4 alkyl and benzyl.

63. A method in accordance with claim 55 in which the R^{32} moieties are each H; and the R^{33} moieties are each independently selected from the group consisting of H and C_1-C_4 alkyl.

64. A method in accordance with claim 55 in which the R^{32} moieties are each H; and the R^{33} moieties are each H.

65. A method in accordance with claim 55 in which R^{34} and R^{35} are each H; y is zero; the R^{32} moieties are each H; and the R^{33} moieties are each H.

5 66. A method in accordance with claim 55 in which R^{34} , R^{35} , R^{36} and R^{37} are each H; y is 1; the R^{32} moieties are each H; and the R^{33} moieties are each H.

10 67. A method in accordance with claim 55 in which R^{34} , R^{35} and R^{36} are each H; R^{37} is OH; y is 1; the R^{32} moieties are each H; and the R^{33} moieties are each H.

15 68. A method in accordance with claim 55 in which R^{34} , R^{35} and R^{36} are each H; R^{37} is NH_2 ; y is 1; the R^{32} moieties are each H; and the R^{33} moieties are each H.

20 69. A method in accordance with claim 55 in which said paramagnetic metal cation is a cation of an element having an atomic number of 22 to 29 or 58 to 70.

70. A method in accordance with claim 55 in which said paramagnetic metal cation is a cation of an element having an atomic number of 24 to 29 or 64 to 68.

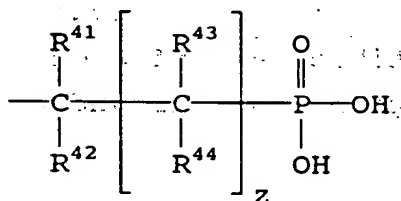
25 71. A method in accordance with claim 55 in which said paramagnetic metal cation is a cation of an element selected from the group consisting of chromium, manganese, iron and gadolinium.

30 72. A method in accordance with claim 55 in which said physiological compatible salt is comprised of said chelate in combination with at least one cation selected from the group consisting of sodium and N-methylglucamine.

35 73. A method in accordance with claim 55 in which said physiologically compatible salt is the pentameglumine

salt of the Gd(III) complex of N,N',N'',N'''-tetrakis-(dihydroxyphosphorylethyl)-1,4,7,10-tetraazacyclononane.

74. A method of preferentially enhancing magnetic resonance image contrast in bone tissue and other tissue of a patient bearing recognition features in common with bone tissue, said method comprising administering to said patient an effective amount of a pharmaceutical agent comprising a physiologically compatible salt of a chelate of a paramagnetic metal cation and a ligand having three or more phosphonate groups of the formula



in which:

R⁴¹, R⁴² and R⁴³ are independently selected from the group consisting of H and alkyl and aryl groups which do not interfere with complexation;

R⁴⁴ is selected from the group consisting of H, OH, NH₂, and alkyl and aryl groups which do not interfere with complexation; and

z is zero or 1.

75. A method in accordance with claim 74 in which said ligand is a cyclic ligand.

76. A method in accordance with claim 74 in which said ligand is a cyclic polyazaalkane comprised of a heterocyclic ring containing ring nitrogen atoms equal in number to said phosphonate groups, with said phosphonate groups bonded to said ring nitrogen atoms.

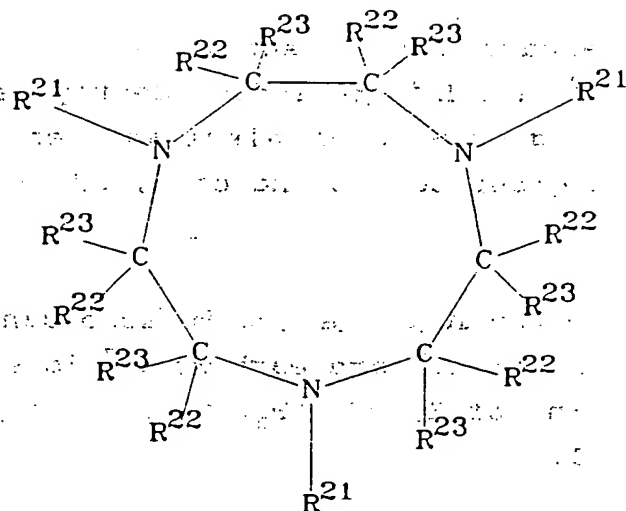
77. A method of preferentially enhancing magnetic resonance image contrast in bone tissue and other tissue of a patient bearing recognition features in common with bone

tissue; said method comprising administering to said patient an effective amount of a pharmaceutical agent comprising a physiologically compatible salt of a chelate of a paramagnetic metal cation and a ligand in which said ligand contains groups which bear specific recognition features for said bone and other tissue.

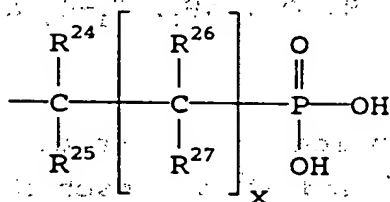
78. A method of preferentially enhancing magnetic resonance image contrast in bone tissue and other tissue of a patient bearing recognition features in common with bone tissue, said method comprising administering to said patient an effective amount of a pharmaceutical agent comprising a physiologically compatible salt of a chelate of a paramagnetic metal cation and a ligand containing phosphonate groups.

79. A method of preferentially enhancing magnetic resonance image contrast in bone tissue and other tissue of a patient bearing recognition features in common with bone tissue, said method comprising administering to said patient an effective amount of a pharmaceutical agent comprising a physiologically compatible salt of a chelate of a paramagnetic metal cation and a ligand containing at least three phosphonate groups.

80. A pharmaceutical agent comprising a chromatographically distinct, physiologically compatible salt of a chelate of a paramagnetic metal cation and a compound having the formula



in which: the R^{21} moieties are each independently



in which R^{24} , R^{25} and R^{26} are independently selected from the group consisting of H and alkyl and aryl groups which do not interfere with complexation; R^{27} is selected from the group consisting of H, OH, NH_2 , and alkyl and aryl groups which do not interfere with complexation; and x is zero or 1;

the R^{22} moieties are each independently selected from the group consisting of H and alkyl and aryl groups which do not interfere with complexation; and the R^{23} moieties are each independently selected from the group consisting of H and alkyl and aryl groups which do not interfere with complexation.

81. A pharmaceutical agent in accordance with claim 80 in which R^{24} , R^{25} and R^{26} are independently selected from the group consisting of H, C_1 - C_8 alkyl, phenyl and benzyl; and R^{27} is selected from the group consisting of H, OH, NH_2 , C_1 - C_8 alkyl, phenyl and benzyl.

82. A pharmaceutical agent in accordance with claim 80 in which R^{24} , R^{25} and R^{26} are independently selected from the group consisting of H, C_1 - C_4 alkyl and benzyl; and R^{27} is selected from the group consisting of H, OH, NH_2 , C_1 - C_4 alkyl and benzyl.

83. A pharmaceutical agent in accordance with claim 80 in which R^{24} , R^{25} and R^{26} are each H; R^{27} is selected from the group consisting of H, OH, NH_2 , C_1 - C_8 alkyl, phenyl and benzyl; and x is 1.

84. A pharmaceutical agent in accordance with claim 80 in which R^{24} , R^{25} and R^{26} are each H; R^{27} is selected from the group consisting of H, OH, NH_2 , C_1 - C_4 alkyl and benzyl; and x is 1.

85. A pharmaceutical agent in accordance with claim 80 in which R^{24} , R^{25} , R^{26} and R^{27} are each H; and x is 1.

86. A pharmaceutical agent in accordance with claim 80 in which R^{24} and R^{25} are independently selected from the group consisting of H, C_1 - C_8 alkyl, phenyl and benzyl; and x is zero.

87. A pharmaceutical agent in accordance with claim 80 in which R^{24} and R^{25} are independently selected from the group consisting of H, C_1 - C_4 alkyl and benzyl; and x is zero.

88. A pharmaceutical agent in accordance with claim 80 in which R^{24} and R^{25} are each H; and x is zero.

89. A pharmaceutical agent in accordance with claim 80 in which the R^{22} and R^{23} moieties are each independently selected from the group consisting of H, C_1 - C_8 alkyl, phenyl and benzyl.

90. A pharmaceutical agent in accordance with claim 80 in which the R^{22} and R^{23} moieties are each independently selected from the group consisting of H, C_1 - C_4 alkyl and benzyl.

91. A pharmaceutical agent in accordance with claim 80 in which the R^{22} moieties are each H; and the R^{23} moieties are each independently selected from the group consisting of H, C_1 - C_8 alkyl, phenyl and benzyl.

92. A pharmaceutical agent in accordance with claim 80 in which the R^{22} moieties are each H; and the R^{23} moieties are each independently selected from the group consisting of H, C_1 - C_4 alkyl and benzyl.

93. A pharmaceutical agent in accordance with claim 80 in which the R^{22} moieties are each H; and the R^{23} moieties are each independently selected from the group consisting of H and C_1 - C_4 alkyl.

94. A pharmaceutical agent in accordance with claim 80 in which the R^{22} moieties are each H; and the R^{23} moieties are each independently selected from the group consisting of H and methyl.

95. A pharmaceutical agent in accordance with claim 80 in which the R^{22} moieties are each H; and the R^{23} moieties are each H.

96. A pharmaceutical agent in accordance with claim 80 in which R^{24} and R^{25} are each H; x is zero; the R^{22} moieties are each H; and the R^{23} moieties are each H.

97. A pharmaceutical agent in accordance with claim 80 in which R^{24} , R^{25} , R^{26} and R^{27} are each H; x is 1; the R^{22} moieties are each H; and the R^{23} moieties are each H.

98. A pharmaceutical agent in accordance with claim 80 in which R^{24} , R^{25} and R^{26} are each H; R^{27} is OH; x is 1; the R^{22} moieties are each H; and the R^{23} moieties are each H.

99. A pharmaceutical agent in accordance with claim 80 in which R^{24} , R^{25} and R^{26} are each H; R^{27} is NH_2 ; x is 1; the R^{22} moieties are each H; and the R^{23} moieties are each H.

100. A pharmaceutical agent in accordance with claim 80 in which said paramagnetic metal cation is a cation of an element having an atomic number of 22 to 29 or 58 to 70.

101. A pharmaceutical agent in accordance with claim 80 in which said paramagnetic metal cation is a cation of an element selected from the group consisting of chromium, manganese, iron and gadolinium.

102. A pharmaceutical agent in accordance with claim 80 in which said physiological compatible salt is comprised of said chelate in combination with at least one cation selected from the group consisting of sodium and N-methylglucamine.

103. A pharmaceutical agent in accordance with claim 80 in which said physiologically compatible salt is the combination of three equivalents of a physiologically compatible cation with the trianionic complex of Fe(III) and N,N',N''-tris(dihydroxyphosphorylmethyl)-1,4,7-triazacyclononane at a pH of about 6.8 to about 7.4.

104. A pharmaceutical agent in accordance with claim 80 in which said physiologically compatible salt is the trisodium salt of the Fe(III) complex of N,N',N''-tris(dihydroxyphosphorylmethyl)-1,4,7-triazacyclononane.

105. A pharmaceutical agent in accordance with claim 80 in which said physiologically compatible salt is the trimeglumine salt of the Fe(III) complex of N,N',N''-tris(dihydroxyphosphorylmethyl)-1,4,7-triazacyclononane.

106. A pharmaceutical agent in accordance with claim 80 in which said physiologically compatible salt is the trisodium salt of the Fe(III) complex of N,N',N''-tris(dihydroxyphosphorylethyl)-1,4,7-triazacyclononane.

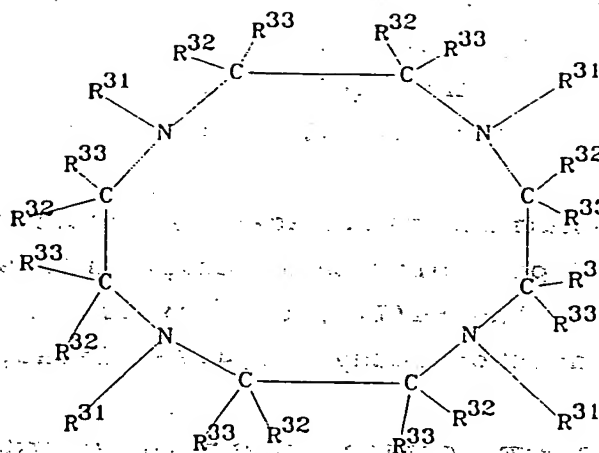
107. A pharmaceutical agent in accordance with claim 80 in which said physiologically compatible salt is the trimeglumine salt of the Cr(III) complex of N,N',N''-tris(dihydroxyphosphorylmethyl)-1,4,7-triazacyclononane.

108. A pharmaceutical agent in accordance with claim 80 in which said physiologically compatible salt is the tetrasodium salt of the Mn(II) complex of N,N',N''-tris(dihydroxyphosphorylmethyl)-1,4,7-triazacyclononane.

109. A pharmaceutical agent in accordance with claim 80 in which said physiologically compatible salt is the trisodium salt of the Mn(III) complex of N,N',N''-tris(dihydroxyphosphorylmethyl)-1,4,7-triazacyclononane.

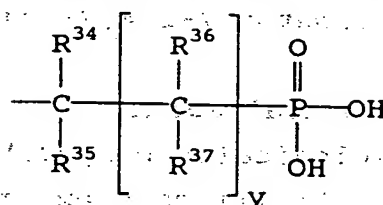
110. A pharmaceutical agent in accordance with claim 80 in which said physiologically compatible salt is the trisodium salt of the Gd(III) complex of N,N',N''-tris(dihydroxyphosphorylmethyl)-1,4,7-triazacyclononane.

111. A pharmaceutical agent comprising a chromatographically distinct, physiologically compatible salt of a chelate of a paramagnetic metal cation and a compound having the formula



in which:

the R³¹ moieties are each independently



in which R³⁴, R³⁵ and R³⁶ are independently selected from the group consisting of H and alkyl and aryl groups which do not interfere with complexation; R³⁷ is selected from the group consisting of H, OH, NH₂, and alkyl and aryl groups which do not interfere with complexation; and y is zero or 1;

the R³² moieties are each independently selected from the group consisting of H and alkyl and aryl groups which do not interfere with complexation; and the R³³ moieties are each independently selected from the group consisting of H and alkyl and aryl groups which do not interfere with complexation.

112. A pharmaceutical agent in accordance with claim 111 in which R³⁴, R³⁵ and R³⁶ are independently selected from the group consisting of H, C₁-C₈ alkyl, phenyl and benzyl; and R³⁷ is selected from the group consisting of H, OH, NH₂, C₁-C₈ alkyl, phenyl and benzyl.

113. A pharmaceutical agent in accordance with claim 111 in which R^{34} , R^{35} and R^{36} are independently selected from the group consisting of H, C_1-C_4 alkyl and benzyl; and R^{37} is selected from the group consisting of H, OH, NH_2 , C_1-C_4 alkyl and benzyl.

114. A pharmaceutical agent in accordance with claim 111 in which R^{34} , R^{35} and R^{36} are each H; R^{37} is selected from the group consisting of H, OH, NH_2 , C_1-C_8 alkyl, phenyl and benzyl; and y is 1.

115. A pharmaceutical agent in accordance with claim 111 in which R^{34} , R^{35} and R^{36} are each H; R^{37} is selected from the group consisting of H, OH, NH_2 , C_1-C_4 alkyl and benzyl; and y is 1.

116. A pharmaceutical agent in accordance with claim 111 in which R^{34} , R^{35} , R^{36} and R^{37} are each H; and y is 1.

117. A pharmaceutical agent in accordance with claim 111 in which R^{34} and R^{35} are independently selected from the group consisting of H, C_1-C_8 alkyl, phenyl and benzyl; and y is zero.

118. A pharmaceutical agent in accordance with claim 111 in which R^{34} and R^{35} are independently selected from the group consisting of H, C_1-C_4 alkyl and benzyl; and y is zero.

119. A pharmaceutical agent in accordance with claim 111 in which R^{34} and R^{35} are each H; and y is zero.

120. A pharmaceutical agent in accordance with claim 111 in which the R^{32} and R^{33} moieties are each independently selected from the group consisting of H, C_1-C_8 alkyl, phenyl and benzyl.

121. A pharmaceutical agent in accordance with claim 111 in which the R^{32} and R^{33} moieties are each independently selected from the group consisting of H, C_1-C_4 alkyl and benzyl.

122. A pharmaceutical agent in accordance with claim 111 in which the R^{32} moieties are each H; and the R^{33} moieties are each independently selected from the group consisting of H, C_1-C_8 alkyl, phenyl and benzyl.

123. A pharmaceutical agent in accordance with claim 111 in which the R^{32} moieties are each H; and the R^{33} moieties are each independently selected from the group consisting of H, C_1-C_4 alkyl and benzyl.

124. A pharmaceutical agent in accordance with claim 111 in which the R^{32} moieties are each H; and the R^{33} moieties are each independently selected from the group consisting of H and C_1-C_4 alkyl.

125. A pharmaceutical agent in accordance with claim 111 in which the R^{32} moieties are each H; and the R^{33} moieties are each independently selected from the group consisting of H and methyl.

126. A pharmaceutical agent in accordance with claim 111 in which the R^{32} moieties are each H; and the R^{33} moieties are each H.

127. A pharmaceutical agent in accordance with claim 111 in which R^{34} and R^{35} are each H; y is zero; the R^{32} moieties are each H; and the R^{33} moieties are each H.

128. A pharmaceutical agent in accordance with claim 111 in which R^{34} , R^{35} , R^{36} and R^{37} are each H; y is 1; the R^{32} moieties are each H; and the R^{33} moieties are each H.

129. A pharmaceutical agent in accordance with claim 111 in which R^{34} , R^{35} and R^{36} are each H; R^{37} is OH; y is 1; the R^{32} moieties are each H; and the R^{33} moieties are each H.

5 130. A pharmaceutical agent in accordance with claim 111 in which R^{34} , R^{35} and R^{36} are each H; R^{37} is NH_2 ; n is 1; the R^{32} moieties are each H; and the R^{33} moieties are each H.

10 131. A pharmaceutical agent in accordance with claim 111 in which said paramagnetic metal cation is a cation of an element selected from the group consisting of chromium, manganese, iron and gadolinium.

15 132. A pharmaceutical agent in accordance with claim 111 in which said physiological compatible salt is comprised of said chelate in combination with at least one cation selected from the group consisting of sodium and N-methylglucamine.

20 133. A pharmaceutical agent in accordance with claim 111 in which said physiologically compatible salt is the pentameglumine salt of the Gd(III) complex of N,N',N'',N'''-tetrakis(dihydroxyphosphorylethyl)-1,4,7,10-tetraazacyclononane.

1. The first part of the paper deals with the general theory of the problem. It is shown that the problem is equivalent to a certain boundary value problem for a second order elliptic equation.

2. In the second part of the paper the author considers the case of a rectangular domain. It is shown that the problem can be reduced to a system of ordinary differential equations.

3. In the third part of the paper the author considers the case of a circular domain. It is shown that the problem can be reduced to a system of ordinary differential equations.

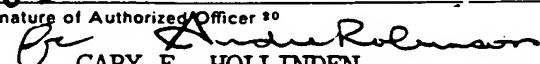
4. In the fourth part of the paper the author considers the case of a domain with a piecewise smooth boundary. It is shown that the problem can be reduced to a system of ordinary differential equations.

5. In the fifth part of the paper the author considers the case of a domain with a smooth boundary. It is shown that the problem can be reduced to a system of ordinary differential equations.

INTERNATIONAL SEARCH REPORT

International Application No

PCT/US90/05967

I. CLASSIFICATION OF SUBJECT MATTER (if several classification symbols apply, indicate all) ³		
According to International Patent Classification (IPC) or to both National Classification and IPC IPC(5): A61B 5/05 A61K 49/00 US CL.: 424/9; 436/173		
II. FIELDS SEARCHED		
Minimum Documentation Searched ⁴		
Classification System	Classification Symbols	
US	424/9, 436/173, 128/653CA 128/653AF	
Documentation Searched other than Minimum Documentation to the Extent that such Documents are Included in the Fields Searched ⁵		
III. DOCUMENTS CONSIDERED TO BE RELEVANT ¹⁴		
Category ⁶	Citation of Document, ¹⁵ with indication, where appropriate, of the relevant passages ¹⁷	Relevant to Claim No. ¹⁸
Y	US, A, 4,804,529 (BARDY et al.) 14 February 1989 See Formula II.	1-133
Y	US, A, 4,880,007 (SADLER et al.) 14 November 1989 See column 2, line 33-column 3, line 20.	1-133
Y	US, A, 4,749,560 (ELGAVISH) 07 June 1988 See column 4, line 20-column 5, line 15.	1-133
Y	US, A, 4,647,447 (GRIES et al.) 03 March 1987 See Formula 16.	1-133
Y	EP, A, 275,215 (SADLER et al.) 20 July 1988 See page 2, line 55-page 3, line 12.	1-133
A	US, A, 4,693,884 (KLEINER et al.) 15 September 1987	
A	US, A, 4,515,766 (ASTRONOVO et al.) 07 May 1985	
<div style="display: flex; justify-content: space-between;"> <div style="width: 45%;"> <p>¹³ Special categories of cited documents:</p> <p>"A" document defining the general state of the art which is not considered to be of particular relevance</p> <p>"E" earlier document but published on or after the international filing date</p> <p>"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)</p> <p>"O" document referring to an oral disclosure, use, exhibition or other means</p> <p>"P" document published prior to the international filing date but later than the priority date claimed</p> </div> <div style="width: 45%;"> <p>"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention</p> <p>"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step</p> <p>"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.</p> <p>"&" document member of the same patent family</p> </div> </div>		
IV. CERTIFICATION		
Date of the Actual Completion of the International Search ²	Date of Mailing of this International Search Report ³	
16 JANUARY 1991	01 MAR 1991	
International Searching Authority ¹	Signature of Authorized Officer ²⁰	
ISA/US	 GARY E. HOLLINDEN	

